

ADVANCES AND APPLICATIONS OF CRISPR-CAS SYSTEMS IN PRECISION GENOME EDITING AND GENETIC DISEASE THERAPY

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ABSTRACT

CRISPR-Cas has transformed the field of gene therapy by providing an unprecedented degree of precision and efficacy with regard to genomic modification over the last decade. These engineered platforms, originally adapted to the adaptive immune system of bacteria, use guide RNA that with the Cas protein can be used to target a particular DNA sequence, which makes it possible to repair mutations, or eliminate diseased genes or insert sequences to cure diseases. Considering such diversity of application, CRISPR has amassed a substantial potential to address a wide range of genetic conditions comprising both monogenetic diseases (e.g., cystic fibrosis and sickle cell anemia) and far more complex conditions (e.g., cancer and infectious diseases). The advent of CRISPR versions and the specific ability to edit the DNA, the Cas9, Cas12, and Cas13 have made the technology even more versatile, allowing disrupting RNA. By its general ability to flex, CRISPR has become a unified system in terms of treating options. However, even with such broad therapeutic application, clinical translation to date is limited by one of two main barriers; the inefficiency of existing delivery methods and the immune reaction against Cas proteins. In the recent past, however, new delivery approaches have increasingly removed these barriers, particularly those based on nanoparticle systems, and optimization of high-fidelity Cas proteins has aimed at improving specificity and safety. In this regard, the field will realize greater efficiencies and effectiveness in employment of this revolutionized technology. This review outlines the mechanisms bases of CRISPR-Cas systems, observes the use of CRISPR-Cas in medical areas in genetic diseases, discusses the current situation of clinical trials progress, and tackles the associated obstacles and ethical issues. It takes into account the current attempts to improve specificity, efficiency, and accessibility as well, highlighting the potential that Crispr has in the development of personalized medicine.

Keywords: CRISPR-Cas systems, gene therapy, genomic editing, genetic disorders, Cas9, personalized medicine, ethical considerations.

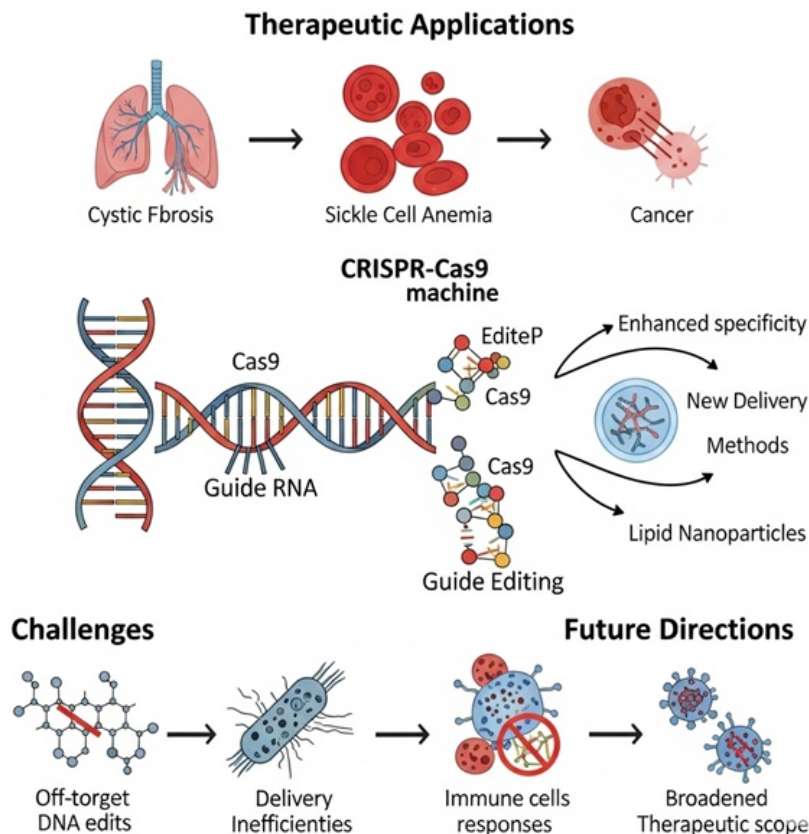


Figure :- CRISPR-Cas Systems: Precision Gene Editing for Therapeutic Innovation

CRISPR-Cas systems revolutionize the sphere of gene therapy because genomic editing becomes simple. This abstract describes the mechanism, and it demonstrates therapeutic uses to a variety of genetic disorders. It also points out the on-going challenges such as off target effect and delivery as well as the future promising directions. Finally, the development of CRISPR will also change personalized medicine.

1. INTRODUCTION

Gene editing is a revolutionary aspect of the biomedical sciences, which have the ability to transform treatment models involving the genetic landscape by manipulating genetic knowledge in terms of sequences [1]. Unlike traditional methods of ameliorating the disease manifestations but at the same time retaining the disease-causing mechanisms, gene-editing approaches aim at repairing or replacing defective alleles that cause the disease to occur [2]. These skills hold the potential to lengthen healing potential across a vast range of hereditary, infectious, and acquired diseases and, as such, provide viable treatment options to ailments that may have previously been considered incurable [3].

Gene editing has promised a lot of treatment in monogenic disorders or disorders caused due to abnormalities in a single gene [4]. As an example, CRISPR technology has undergone an intensive study into sickle cell anemia and cystic fibrosis as therapeutic treatment [5]. CRISPR potentially can help to cure the affected cells by repairing the genetic issue that gives rise to it [6]. On the same note, preclinical studies that use gene editing technologies to reinstate proteins have demonstrated potential in conditions like Duchenne [7].

The treatment of more complex disorders beyond monogenic diseases can be deployed through gene editing. CRISPR technology is the technology that is being used to edit cancer immune cells [8]. As an example, CRISPR-engineered T cells (CARs) have successfully been used to treat leukaemia and lymphoma patients because the T cells can disrupt the specific antigens present on the cancerous cells [9]. Furthermore, the possibility to edit genomes can allow to deactivate oncogenes or re-activate tumor suppressor genes

and this is one of the direct ways to intervene with the process of cancer development [10]. The field of gene editing is also applied to the treatment of infectious diseases. It attacks the genomes of pathogen [11]. CRISPR systems can interfere with their cycle of replication. New developments in the gene-editing technology most prominently include CRISPR-Cas13, which has broadened the horizons of therapeutics by allowing targeting of viruses like HIV and SARS-Co V-2 at a level as specific as RNA, thereby creating an entirely new concept of antiviral intervention [12]. Parallel to this, the possibility of controlling antibiotic resistance has become a topic of interest as well. Disruption of gene resistance or the enhancement of drug-susceptibility to currently available drugs can be the appropriate mode of countering bothersome diseases such as *Clostridioides difficile* colitis or methicillin-resistant *Staphylococcus aureus* [13]. In addition to treatment of clinical problems, preventive opportunities are opened when germline editing is used and then the genetic material of the embryo can be altered to prevent the onset of inherited disorders like Huntington disease, Tay-Sachs disease or thalassemia [14]. Besides, the development of new delivery agents such as cell-penetrating peptides, viral vectors, and nanoparticles formed of lipids has solved issues concerning the delivery of gene editing instruments to the diseased tissue or cells [15]. Any of these developments are making the idea of personalized medicine a reality where medicine can be tailored to the genetics of an individual to optimize a therapy outcome. The practice of gene editing is still nonetheless confronted with a number of obstructions when applied to the medical field [16]. The most noteworthy among them is off-target effects possibility, immune responses to Cas proteins and challenges in addressing desired tissues/ organs with the editing tools [17]. Ethical issues of germline editing, fair distribution of gene therapies, and possibilities of misuse of such technologies should also be brought up. Nonetheless, research and development in this field are still broadening the horizons of gene editing uses, with rare genetic disorder treatments getting coverage to more extensive socially significant problems such as cancer and infectious diseases [18]. The evaluation of CRISPR-cas system is shown in Table 1.

Table 1:- Evolution of CRISPR-Cas Systems

CRISPR Tool	Target	Unique Features	PAM Requirement	Applications	Reference
Cas9	DNA	Double-strand breaks, requires tracrRNA	NGG	Gene editing, knockout models, HDR-based repair	[19]
Cas12a (Cpf1)	DNA	Staggered cuts, simpler guide RNA, multiplexing	TTTV	Diagnostic platforms, plant editing	[20]
Cas13	RNA	RNA editing, collateral cleavage	None	RNA knockdown, virus detection (e.g., SHERLOCK)	[21]
Cas14	ssDNA	Ultra-small,	None or flexible	SNV detection	[22]
		high specificity		diagnostics	
Base Editors	DNA	Converts A→G or C→T without DSBs	Cas9-based	Precision mutation correction in monogenic diseases	[23]
Prime Editors	DNA	Versatile insertions/deletions/base changes	Cas9 nickase-based	Gene therapy, disease modeling, safer genome edits	[24]

2. CRISPR-CAS SYSTEMS: MECHANISM OF ACTION

It has revolutionized genome where it provides a very precise, and convenient tool of changing the genetic material, and these systems origins germinate on the bacterium and archaeal adaptive immune systems that were found in contrast to overcome invading viruses by identifying and cleaving the DNA or RNA of the virus. This natural defence mechanism has been shaped into a powerful modifying genes tool which is applicable in modifying certain genomic regions [25].

Figure 1 illustrates the three important steps in the process of the CRISPR-Cas systems, and they are, recognition, cleavage, and repair.

Step 1: Recognition

The specification uses the design to initiate the whole process and entry of a guide ribonucleic acid (gRNA) into the cell. The gRNA seems to consist of two major parts: the scaffold conformation that binds to the a sequence which is complementary to the target DNA, ts, and the Cas protein [26]. The system recognizes the target through sequence complementarity between the gRNA and the (PAM) that is

near the target site. PAM is selective and reduces the chance of off-target effects since it is used to bind to the Deoxyribo nucleic acid on behalf of Cas protein [27].

Step 2: Cleavage

When the Cas protein has been guided towards the destination. Deoxyribo nucleic acid, in the target site. Cas enzymes like Cas9 are molecular shears that cut down the DNA [17]. This is possible due to the capability of the protein. There are various types of Cas proteins and these proteins cut in different ways. To give an example, Cas9 makes blunt end cuts and other Cas variants such as Cas12 and Cas 13 have different pattern cleavages, targeting not only DNA, but also single-strands and RNA molecules [28].

Step 3: Repair

The last process presupposes the natural repair of cell, which repairs the DNA break created upon the Cas protein insertion. The situation has two major repair paths:

Non-Homologous End Joining (NHEJ): It is a repair process, in which the fragments of the chromosome DNA directly connected without a homologous template. It makes it ideal when it comes to gene knockouts as it tends to induce minor

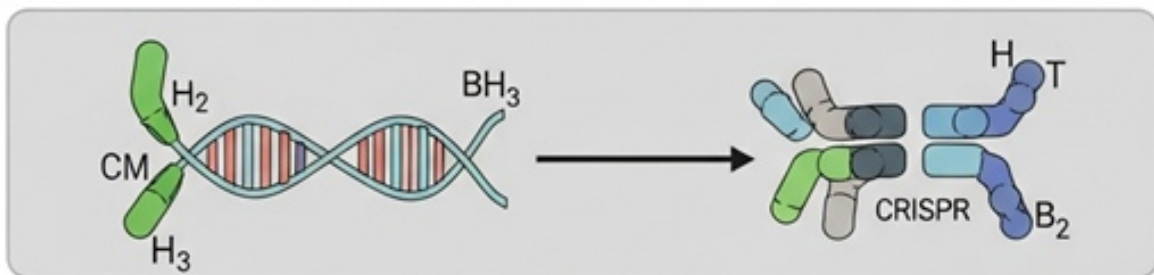
additions or subtractions (indels) [29].

Homology-Directed Repair (HDR): A homologous DNA is used as a template of this cascade to repair the break properly. HDR is the preferred mode of controlling the fine-tuned changes of the gene, like fixing a mutation or adding a new gene [30]. But in comparison to NHEJ, HDR is not as effective and usually the cell must be in certain stages of the cell cycle [31].

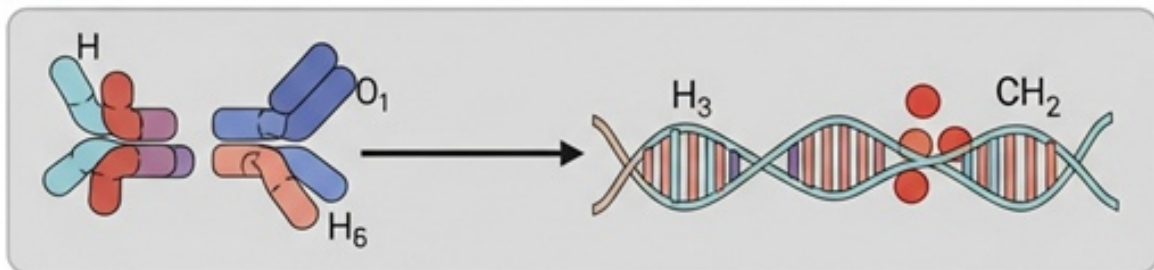
These three processes attack recognition, cleavage and repair- all these together to render these systems an effective genome editing tool. This variable is the repair mechanism that considerably affects the gene-editing process through which the researchers can customize the system to suit a particular need [1].

CRISPR-Cas Systems

Step 1: Recognition



Step 2: Cleavage



Step 3: Repair

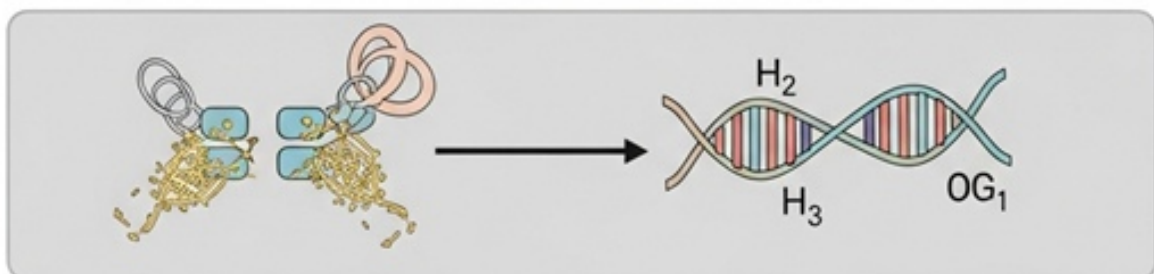


Figure 1:- Mechanism of CRISPR-Cas Systems

The illustration shows key steps of operation of CRISPR-Cas systems: recognition of the target DNA chain by guiding RNA (RNA-guided), accurate cleavage of double-stranded DNA by the Cas protein, and the repair of a cell by one of the modes of the HDR and NHEJ repair mechanisms. This kind of representation represents the complexity of a CRISPR-Cas system as a gene-editing system.

2.1. CRISPR Variants:-

1. Type II (Cas9): Cas9 is also the most commonly applied protein both in rese applications and in clinical practice, as it proves very precise and flexible. It has been used to make gene knockouts to understand the qualities of a gene, to rectify genetic rearrangements of monogenic diseases such as HbSS disease, to alter immune cells as a cancer treatment, and to

create genome editing [32].

2. Type V (Cas12): Cas12 systems have been marvelous especially in diagnostics and anti-viral therapies. This is the case with viral pathogens which have been detected using the Cas12 proteins integrated into such platforms as (Specific High-sensitivity Enzymatic Reporter unLOCKing) [33].

3. Type VI (Cas13): Cas13 has RNA editing as its specialty, thus it is applicable in gene silencing and in fighting RNA viruses. This system has been used in the design of diagnostic reagents of infectious diseases and may have potential in the treatment of RNA-based afflictions such as Huntington disease [34]. Table 2 illustrates the CRISPR Variants, their characteristics, and uses thereof.

Table 2:- CRISPR Variants and their applications

Type	Cas Protein	Features	Applications	Refrences
Type II	Cas9	Most widely used; introduces double-strand breaks with high precision	Gene knockout, correction of point mutations, therapeutic gene editing	[35]
Type V	Cas12	Capable of cutting both DNA and RNA; has collateral activity	Applications in anti-viral therapies, DNA/RNA targeting, diagnostics	[36]
Type VI	Cas13	RNA-specific editing without affecting DNA; introduces single-strand breaks	Gene silencing, RNA diagnostics, treating RNA viruses such as SARS-CoV-2 and HIV	[37]

3. APPLICATIONS IN DISEASE TREATMENT

The CRISPR technology has become a management tool to control various pathological conditions by the direct targeting and modification of genetic basis of conditions [38]. It has the potential to provide highly accurate genetic modification, which has provided new possibilities in the treatment process, especially among disorders that include a certain established genetic factor [39]. Some of the examples of the usage of CRISPR in disease treatment are presented below and divided into monogenic diseases, cancer treatment, and the treatment of infectious diseases.

3.1 Monogenic Disorders

Monogenic diseases which in most cases are the result of mutations that take place in a single gene are perfectly suitable targets of CRISPR-based therapeutic interventions. CRISPR will be able to identify the defective gene and by correcting the mutation and replacing it by a functional gene, be able to offer a possible permanent cure [40].

1. Sickle cell anemia: The recent advances in the development of gene editing technology have focused on its work precision, efficiency, and safety [35]. The process of CRISPR-based treatments includes the treatment of hematopoietic stem cells (HSCs) so that the HBB mutation can either be fixed or fetal hemoglobin (HbF) can again be produced through the activation of regulatory genes such as BCL11A. These modified HSCs are reintroduced to the patient to create healthy red blood cells, and this gives the patient a functional cure to this disease [41]. Cases of success of giving clinical trials have proven that the patients get to reach higher levels of sustaining a normal hemoglobin level, in the long term.

2. Cystic Fibrosis: Cystic fibrosis is caused by mutations in the CFTR gene, which encodes a protein involved in chloride ion transporter [42]. In epithelial this technology has been used to treat mutations in CFTR genes in patient epithelial cells resulting in normal protein function [43]. This methodology is promising at the preclinical level, and current research

involves how to best deliver the method to the lung tissue through in vivo models. In its initial stages, CRISPR has the potential to cure cystic fibrosis at its genetic level.

3. Duchenne Muscular Dystrophy (DMD): DMD is characterized by the lack of preserved dystrophin protein that enables intact muscles, as it is the case with DMD because of the mutations that occur in DMD gene [44]. CRISPR-based technologies have proven to correct dystrophin expression in preclinical models by either removing or repairing mutated areas of the gene [45]. In animal models, it has been demonstrated that the effectiveness of muscle performance and the progress of the disease slows down indicating the potential of human trials [46].

3.2 Cancer Therapy

The most studious concentration of CRISPR has been on cancer as it is a multifactorial disease caused by genetic and epigenetic alterations. The technology is finding application in improving the efficacy of current treatments and coming up with new ones [47].

1. Knock out of Oncogenes: CRISPR can inactivate oncogenes, which are cancer growth promoters by adding specific mutations [48]. As an illustration, members of an oncogenic gene cluster such as KRAS and MYC that are typically mutated in most cancers have been disrupted using CRISPR. The measure minimizes the growth potential of the cancer cells and makes them more vulnerable to other treatments [49].

2. Improving the efficacy of Immune Cells: CRISPR has been used to create Chimeric Antigen Receptor (CAR) T-cells where T-cells have been genetically modified so that they are receptive to the presence of cancerous cells and destroy them [50]. CRISPR can also enhance the outcome of CAR-T treatment by removing immune checkpoint genes such as PD-1 and CTLA-4 that inhibit the immune system [51]. Such adjustments enable T-cells to be functional at a longer duration and this enhances their sensitivity to identify and destroy tumours. CRISPR-improved CAR-T therapies have already been approved in some malignancies including leukaemia and lymphoma [1].

3. Tumor Suppressor Gene Activation: It is also possible to

reactivate tumor suppressor genes which are silenced in cancer cells using CRISPR. CRISPR re-activates these genes by editing the regulatory regions of the gene thereby preventing the development of tumor [52].

3.3 Infectious Diseases

CRISPR has significantly helped in fighting infectious diseases by focusing on the genetic material of a pathogen, such as a virus, bacteria, and a parasite [53].

1. Degradation of Viral Genomes: Viral RNA can also be degraded by CRISPR systems targeting RNA, most commonly using Cas13 due to its small size and simplicity. By directing Cas13 to target viral RNA, one can use it to disrupt the reproducing virus, offering a new therapeutic treatment against RNA viruses like the human immunodeficiency virus (HIV) the SARS-CoV-2 virus [54]. For instance, Cas13 has been applied to disrupt the reproducing SARS-CoV-2 virus, indicating promise. Closely related strategies are under development against HIV, in which CRISPR is to be used to cleave the proviral DNA integrated into a host genome [55].

2. Bacterial Infections: Antibiotics resistance becomes a worldwide issue. Biological weapon CRISPR has the ability to break into the bacteria genomes and render resistance genes, which leaves the bacteria vulnerable to current antibiotics [56]. Also, it is possible to use CRISPR to kill bacteria directly by targeting necessary genes, providing high specificity in the fight against difficult-to-treat infections without damaging useful microbiomes [57].

3. Parasites: CRISPR has undergone preclinical application to affect and edit the genome of parasitic organism including Plasmodium (which causes malaria). CRISPR has the potential to control parasitic diseases in a new way by interrupting genes necessary to survival or transmission of parasites [58].

The flexibility of CRISPR is still increasing its clinical promising, which gives hope to the inability of treating some of the diseases [32]. Although several challenges should still be addressed, including delivery and ethical issues, the blistering development of the technology leads to the fact that its influence on medicine will be massive [59]. Table 3 demonstrates the uses of CRISPR in the therapeutic process of diseases.

Disease	Gene Target	Outcome	Clinical Trial Status	References
Sickle Cell Anemia	HBB	Restored hemoglobin production	Ongoing (e.g., CTX001 trial)	[60]
Cystic Fibrosis	CFTR	Corrected chloride channel function	Preclinical studies	[61]
Duchenne Muscular	DMD	Restored dystrophin expression	Preclinical (animal models)	[7]
Cancer (CAR-T Therapy)	PD-1, CTLA-4	Enhanced T-cell activity	Approved for certain cancers	[62]
HIV	Proviral DNA	Disrupted viral replication	Preclinical (ongoing research)	[63]
SARS-CoV-2	Viral RNA	Inhibited viral replication	Experimental (Cas13-based tools)	[64]

4. CHALLENGES

Although it has power to transform, CRISPR technology has quite a number of problems that need to be resolved to make it safe, effective, and applicable in diverse areas. Issues such as off-target effects, delivery mechanism and the immunogenicity are the most prominent challenges of these substances [65]. Each of them has distinct challenges to clinical use of CRISPR-based therapeutics and calls for creative solutions. Table 4 & Figure 5 presents the Current Challenges in CRISPR-Cas Therapy.

4.1 Off-Target Effects

Off-target effects are the result of CRISPR system bringing about unwanted genomic editions in places that are not destined. These malfunctions may have unpredictable results that can be disruption of genes, activation of Oncogenes or inactivation of tumor suppressor genes which may result in oncogenesis or other fatal complications [66].

Mechanism: When some mismatches occur on the guide Ribo Nucleic acid (gRNA) sequences and the non-target Deoxyribo Nucleic acid sequences, the Cas protein cuts the wrong sites which is known as off-target effects. Such off-target mutations undermine the precision and security of the CRISPR based treatments [67].

4.2 Delivery Mechanisms

The treatment of gene-editing uses the introduction of CRISPR machinery within its target tissues and cells. The key

barriers are surmounting biological barriers, reaching cellular specificity, and making CRISPR components stay there and be expressed over time without inducing toxicity [68].

Mechanism: This predicament of delivery that confronts synthetic Cas proteins and related gRNA constructs further receives a significant setback to its low molecular weight and the need of specific intracellular delivery. The classic vehicles, i.e., mostly viral vectors, are also subject to limitations, in particular, immunogenic activation and low loading capacity [69].

4.3 Immunogenicity

Immunogenicity of CRISPR elements, especially the Cas proteins of bacterial origin, may cause immune responses to the host. Such immune reactions can diminish the effectiveness of the treatment and create a threat of adverse reactions [70].

• **Mechanism:** Cas proteins Cas proteins, including SpCas9 of *Streptococcus pyogenes*, are perceived by immune system as foreign antigens. This identification spurs the immune responses that break down the components of CRISPR or result in inflammatory responses. This is worsened by pre-existing immunity to Cas proteins that is brought about by pre-exposure of the source bacteria [71].

Table 4: Current Challenges in CRISPR-Cas Therapy

Challenge	Description	Proposed Solutions	References
Off-Target Effects	Unintended gene edits causing adverse effects	Improved gRNA design, high-fidelity Cas proteins, and validation assays	[32]
Delivery Mechanis	Poor delivery to target tissues and cells	Nanoparticle-based delivery, AAV vectors, electroporation	[72]
Immune Responses	Host immunity to bacterial Cas proteins	Engineering less immunogenic Cas proteins, temporary immunosuppression, humanized systems	[68]

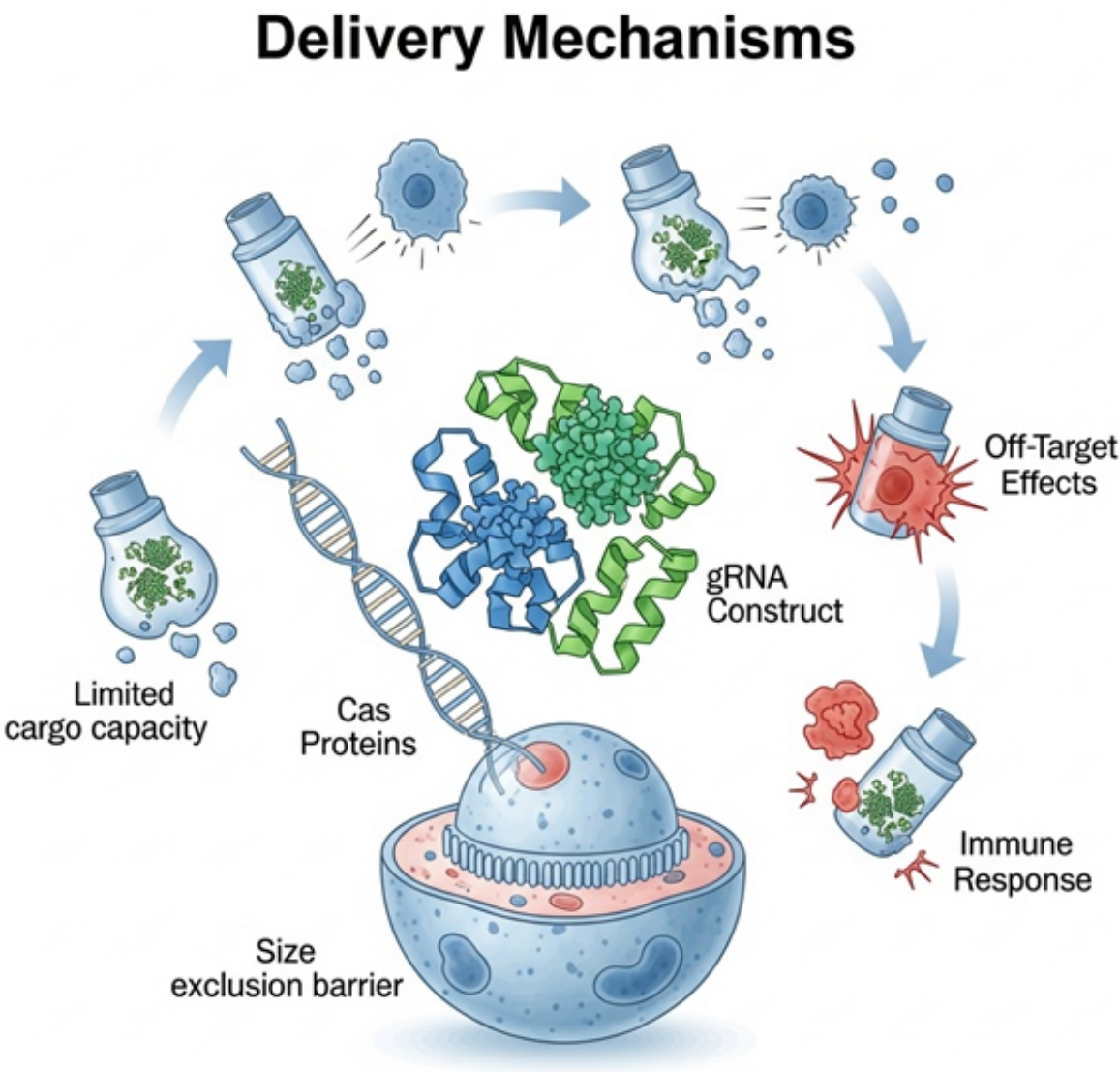


Figure 2 :Challenges in Delivering CRISPR Components for Gene Editing

The challenge in gene editing represented in this image is the Delivery Mechanisms. It reveals the challenges of getting big machinery of CRISPR (Cas proteins and gRNA constructs) into target cells because of size exclusion and biological obstructions. It also addresses the drawbacks of conventional viral vectors such as limited capacity to carry cargoes, off-target effect, and triggering of immune responses.

.ETHICAL CONSIDERATIONS

The evolution of CRISPR-Cas-technology has opened up new opportunities that are unprecedented in terms of the possibility of treatment and prevention of diseases. Nevertheless, the usage of it gives rise to some very important ethical considerations that must be critically discussed [39]. These issues include what these somatic and germline editing solutions may imply, the problem of fair access to treatments and the challenges of a patchwork system of regulation across different countries. It is crucial to contend with such considerations to make the technology responsibly and fairly used [73].

5.1 Somatic vs. Germline Editing

The discussion of the difference between germline and somatic editing is one of the most discussed ethical topics of CRISPR.

Somatic Editing: Somatic cell editing genetic modifications are designed to affect only the individual treated where the editing will not be passed onto the future generations since it is done on cells that are non-reproductive [74]. This method is traditionally discussed as ethically permissible especially when it comes to treatment of severe or life-threatening diseases like HBSS or Duchenne dystrophy. It has the prospect of accuracy treatments with minimal societal effects in the long run [75].

Germline Editing: Examples of germline editing include editing the DNA of eggs or sperm or editing of the DNA of the embryo, which also puts forth heritable changes to the offspring [76]. Although the concept of germline editing has potential to eliminate genetic disorders within a family tree, it is also very ethically disconcerting:

1. **Unintended Consequences:** Some adverse effects might be due to off-target or unexpected genetic interactions that may have irreversible effects on future generations [1].
2. **Moral Implications:** Critics are more eager to claim that germline editing is an ethical line that should not be crossed since it inhibits the evolutionary process of a human being as it is [77].

3. **Designer Babies:** Misapplication of the possibility of creating superior humans with such traits as superior intelligence or athletic ability could create unequal advancements in people as well as insurgent questions as to genetics [78].

5.2 Equity in Access

Although CRISPR-based therapeutics have passed the test of hope when it comes to addressing genetic diseases, they are expensive and incredibly technically demanding, which does not contribute to equitable access, especially in resource-poor countries [79].

Economic Disparities: There is a huge financial input that is needed in the development CRISPR therapies, its testing and application and such treatments end up being too expensive even to people in the developing world [16]. As an extreme example, the cost to treat potential patients with a therapy such as CRISPR-based interventions to treat sickle cell is currently estimated at \$1000s per patient [80].

- **Global Healthcare Inequities:** CRISPR-based therapies are only effective when used on an advanced healthcare facility, and since these advances are only available in developed countries that have well established healthcare systems, it can only be used to treat the wealthier countries. Such discrepancy will vastly increase, leading to the widening of the health gap across the world [81].

5.3 Regulatory Frameworks

The existence of many different standards in terms of CRISPR-based therapies complicates their ethical and clinical usage [59].

Lack of Uniformity:

At one extreme, some states at least two, Somalia and the United Arab Emirates have developed blanket bans on germline interventions, a virtual ban on all research activities implying manipulation of reproductive cells. On the other end the spectrum, a few countries, most prominently China and Russia, have already approved of clinical studies involving germline editing, and, thus, allowed the human genome to be directly and naturally altered. Such institutional divide creates a lot of regulatory uncertainty and increases public uncertainty about gene-editing tourism, the

phenomenon, where tourism clients want to use unregulated or under-regulated procedures in other countries [82].

Safety standards:-

The regulation of safety levels in the preclinical and clinical stage of the experiment is inevitable to eliminate the possibility of unpredictable consequences and to clarify whether gene-editing treatment could cause excessive harm to the patients. Without a global agreement framework, the process of assessing whether the research meets ethical and safety standards turns problematic, and, therefore, inaccurate [83].

FUTURE DIRECTIONS

The favorable tendencies of the CRISPR technology are expected to overcome the existing limitations and unlock new perspectives of its use. In-depth focus on increasing the degree of specificity of genome-editing systems has resulted in significantly more off-target activity-buffered targeted nucleases like SuperFi-Cas9 and Cas12b. These nucleases have smaller tolerances of protospacer-adjacent sequence motifs, report low global genomic hits, and do not lose a lot of in vivo efficiency. The enhanced accuracy makes gene-based treatments more secure and reliable. Finally, the CRISPR toolkit is still expanding due to the Cas12c and high-fidelity Cas9 variants, strengthening the genome editing potential with most cell and organismal lineages. In the same way, the editing power of RNA of CRISPR-Cas13 has different benefits, especially the capability to bring about a transient and even reversible gene editing and selectively blocks the build-up of mRNA implicated in diseases like retinal conditions. In the biomedical field, more than thirty clinical trials using CRISPR-edited T cells in cancer patients have already been started, with considerable advancements made on the allogeneic CAR-T structures- an advance that might make it possible to create off-the-shelf immune cell therapies. It is also by CRISPR that combination approaches are now being integrated: combining with RNA interference, or single-cell omics, these ways are providing more sturdy multi-dimensional therapeutics against complex diseases. areas of applications are still expanding to diagnostics (a quick pathogen test), sustainable manufacturing (engineered pathogens of bioproduction), agriculture (precision crop

engineering), and ecological engineering (gene drives and species recovery). All such improvements are outside of the scope of traditional genome editing, with potential revolutionary effects in biomedicine, biotechnology, and in environmental sustainability.

7. CONCLUSION

CRISPR-Cas system is introducing a new era of gene therapy with accurate and efficient ways of treatment of many genetic diseases, cancers, and viral diseases. CRISPR-Cas has become a powerful technology that can potentially address genetic abnormalities by correcting them through specific genome editing; however, a range of concerns, including efficient delivery mechanisms, undesirable activations of immune system responses, and ethical status of germline editing, are still up to be scrutinized with specific severity. The world today has been striving to take safety, accessibility, and equity to greater heights; one of the approaches is the new vehicle of delivering such a drug or vaccine, this has been introduced through the use of lipid nanoparticle and the viral vectors with outdated immune reaction. At the same time, researchers are tightening control over factors of both time and space of gene editing to minimize off-target effects. The development of ethical frameworks is becoming increasingly fast in order to address the growing concerns since scientific innovation and social values deserve much attention, particularly focusing on the heritable genome modification. Increased international cooperation and interaction of the population should be essential to guarantee equal access and avoid inequality in access to CRISPR technology-based medicines. By virtue of persisted research, open regulatory regulation, and interdisciplinary cooperation, CRISPR has a transformative potential to relaunch clue medication, by enabling patent-specific therapies that extend patient outcome, save certain medical expenses, and eventually place healthcare more accurate, effective, and available all over the globe.

CONFLICT OF INTEREST

None

FUNDING

None

REFERENCES:-

1. Srivastav, A. K., Mishra, M. K., Lillard Jr, J. W., & Singh, R. (2025). Transforming pharmacogenomics and CRISPR gene editing with the power of artificial intelligence for precision medicine. *Pharmaceutics*, 17(5), 555.
2. Qie, B., Tuo, J., Chen, F., Ding, H., & Lyu, L. (2025). Gene therapy for genetic diseases: challenges and future directions. *MedComm*, 6(2), e70091.
3. Levesque, S., & Bauer, D. E. (2025). CRISPR-based therapeutic genome editing for inherited blood disorders. *Nature Reviews Drug Discovery*, 1-19.
4. Khawaja, S., Ali, R. H., Ahmed, I., & Umair, M. (2025). Gene Therapy in Rare Genetic Disorders: Current Progress and Future Perspectives. *Current Genomics*.
5. Ahmed, R., Alghamdi, W. N., Alharbi, F. R., Alatawi, H. D., Alenezi, K. M., Alanazi, T. F., & Elsherbiny, N. M. (2025). CRISPR/Cas9 system as a promising therapy in Thalassemia and Sick Cell Disease: a systematic review of clinical trials. *Molecular Biotechnology*, 1-10.
6. Azani, A., Sharafi, M., Doachi, R., Akbarzadeh, S., Lorestani, P., Haji Kamanaj Olia, A., ... & Behfar, Q. (2025). Applications of CRISPR-Cas9 in mitigating cellular senescence and age-related disease progression. *Clinical and Experimental Medicine*, 25(1), 1-21.
7. Haque, U. S., & Yokota, T. (2025). Gene Editing for Duchenne Muscular Dystrophy: From Experimental Models to Emerging Therapies. *Degenerative Neurological and Neuromuscular Disease*, 17-40.
8. Youssef, E., Fletcher, B., & Palmer, D. (2025). Enhancing precision in cancer treatment: the role of gene therapy and immune modulation in oncology. *Frontiers in Medicine*, 11, 1527600.
9. Kamli, H., & Khan, N. U. (2025). Revolutionising cancer intervention: the repercussions of CAR-T cell therapy on modern oncology practices. *Medical Oncology*, 42(7), 228
10. Akhter, S., Ansari, M. A. S., Tiva, M. G., & Bhuyian, M. S. (2025). Improving Treatments for Oral Diseases, Head and Neck Cancers, as well as Developing New Technologies. *Pathfinder of Research*, 3(1), 1-25.
11. Manzoor, S., Nabi, S. U., Rather, T. R., Gani, G., Mir, Z. A., Wani, A. W., ... & Manzar, N. (2024). Advancing crop disease resistance through genome editing: a promising approach for enhancing agricultural production. *Frontiers in genome editing*, 6, 1399051.
12. Liu, L., & Pei, D. S. (2022). Insights gained from RNA editing targeted by the CRISPR-Cas13 family. *International Journal of Molecular Sciences*, 23(19), 11400.
13. Gopikrishnan, M., Haryini, S., & C, G. P. D. (2024). Emerging strategies and therapeutic innovations for combating drug resistance in *Staphylococcus aureus* strains: A comprehensive review. *Journal of Basic Microbiology*, 64(5), 2300579.
14. Wong, P. K., Cheah, F. C., Syafruddin, S. E., Mohtar, M. A., Azmi, N., Ng, P. Y., & Chua, E. W. (2021). CRISPR gene-editing models geared toward therapy for hereditary and developmental neurological disorders. *Frontiers in Pediatrics*, 9, 592571.
15. Zu, H., & Gao, D. (2021). Non-viral vectors in gene therapy: recent development, challenges, and prospects. *The AAPS journal*, 23(4), 78.
16. Olaghere, J., Williams, D. A., Farrar, J., Büning, H., Calhoun, C., Ho, T., ... & Reagan-Udall Foundation for the FDA. (2025). Scientific Advancements in Gene Therapies: Opportunities for Global Regulatory Convergence.
17. Baig, M. M. F. A., & Chien, W. T. (2025). Nanotechnological Approaches for the Targeted Delivery of CRISPR-Cas Systems for Genomic Modifications, Biomolecular Sensing, and Precision Medicine. *Biomaterials Science*.
18. Gibelli, F., Ricci, G., & Bailo, P. (2025). Genome Editing in Medicine: A Scoping Review of Ethical, Bioethical, and Medico-Legal Implications. *Journal of Law, Medicine & Ethics*, 1-9.
19. Skryabin, B. V., Braun, D. A., Kaiser, H., Gubar, L., Seeger, B., Khanam, T., ... & Rozhdestvensky, T. S. (2025). CRISPR-Cas9 HDR Optimization: RAD52, Denatured and 5'-Modified DNA Templates in Knock-In Mice Generation. *bioRxiv*, 2025-05.
20. Karimi, M., Ghorbani, A., Niazi, A., Rostami, M., & Tahmasebi, A. (2025). CRISPR-Cas13a as a next-generation tool for rapid and precise plant RNA virus diagnostics. *Plant Methods*, 21(1), 83.
21. Chen, L., Duan, L., Li, J., Chen, J., Liao, D., He, N., ... & Hu, Z. (2025). Advances in CRISPR-based gene editing technology and its application in nucleic acid detection. *Biocell*, 49(1), 21.
22. Son, H. (2024). Harnessing CRISPR/Cas systems for DNA and RNA detection: principles, techniques, and challenges. *Biosensors*, 14(10), 460.
23. Reshetnikov, V. V., Chirinskaite, A. V., Sopova, J. V., Ivanov, R. A., & Leonova, E. I. (2022). Translational potential of base-

- editing tools for gene therapy of monogenic diseases. *Frontiers in Bioengineering and Biotechnology*, 10, 942440.
24. Lu, Y., Happi Mbakam, C., Song, B., Bendavid, E., & Tremblay, J. P. (2022). Improvements of nuclease and nickase gene modification techniques for the treatment of genetic diseases. *Frontiers in Genome Editing*, 4, 892769.
25. Navarro, C., Díaz, M. P., Duran, P., Castro, A., Díaz, A., Cano, C., ... & Bermúdez, V. (2025). CRISPR-Cas Systems: A Functional Perspective and Innovations. *International Journal of Molecular Sciences*, 26(8), 3645.
26. Omura, S. N., Alfonse, L. E., Ornstein, A., Morinaga, H., Hirano, H., Itoh, Y., ... & Nureki, O. (2025). Structural basis for target DNA cleavage and guide RNA processing by CRISPR-Cas?2. *Communications Biology*, 8(1), 876.
27. Modrzejewski, D., Hartung, F., Lehnert, H., Sprink, T., Kohl, C., Keilwagen, J., & Wilhelm, R. (2020). Which factors affect the occurrence of off-target effects caused by the use of CRISPR/Cas: a systematic review in plants. *Frontiers in plant science*, 11, 574959.
28. Alam, Q., Rafeeq, M., & Khan, M. U. (Eds.). (2025). *Innovations and Implications in Molecular Diagnostics*. CRC Press.
29. Ilioaia, O., Dudragne, L., Brocas, C., Meneu, L., Koszul, R., Dubrana, K., & Xu, Z. (2025). The CST complex mediates a post-resection non-homologous end joining repair pathway and promotes local deletions in *Saccharomyces cerevisiae*. *Cell Genomics*.
30. Haider, S., & Mussolino, C. (2025). Fine-Tuning Homology-Directed Repair (HDR) for Precision Genome Editing: Current Strategies and Future Directions. *International Journal of Molecular Sciences*, 26(9), 4067.
31. Jin, Y. Y., Zhang, P., & Liu, D. P. (2025). Optimizing homology-directed repair for gene editing: the potential of single-stranded DNA donors. *Trends in Genetics*.
32. Alariqi, M., Ramadan, M., Yu, L., Hui, F., Hussain, A., Zhou, X., ... & Jin, S. (2025). Enhancing Specificity, Precision, Accessibility, Flexibility, and Safety to Overcome Traditional CRISPR/Cas Editing Challenges and Shape Future Innovations. *Advanced Science*, e2416331.
33. Smailov, S., Gabdullina, E., Lessova, Z., & Assembayeva, E. (2025). Gene editing by CRISPR-Cas-biotechnological applications. *Fundamental and Experimental Biology*, 11730(1), 65-73.
34. Allemailem, K. S., Rahmani, A. H., Almansour, N. M., Aldakheel, F. M., Albalawi, G. M., Albalawi, G. M., & Khan, A. A. (2025). Current updates on the structural and functional aspects of the CRISPR/Cas13 system for RNA targeting and editing: A next generation tool for cancer management. *International Journal of Oncology*, 66(5), 1-21.
35. Zang, S. S., Zhang, R., Zhang, J. R., Zhang, X., & Li, J. (2025). Progress, Applications and Prospects of CRISPR-Based Genome Editing Technology in Gene Therapy for Cancer and Sickle Cell Disease. *Human Gene Therapy*, 36(11-12), 858-869.
36. Xiang, W., Lin, X., Yang, Y., Huang, L., Chen, Y., Chen, J., & Liu, L. (2025). Cas12h is a crRNA-guided DNA nickase that can be utilized for precise gene editing. *Cell Reports*, 44(5).
37. Zhou, S., Yang, S., Xu, J., & Zhu, G. (2025). Engineering circular guide RNA and CRISPR-Cas13d-encoding mRNA for the RNA editing of Adar1 in triple-negative breast cancer immunotherapy. *bioRxiv*, 2025-07.
38. Shahid, A., Zahra, A., Aslam, S., Shamim, A., Ali, W. R., Aslam, B., ... & Arshad, M. I. (2025). Appraisal of CRISPR Technology as an Innovative Screening to Therapeutic Toolkit for Genetic Disorders. *Molecular Biotechnology*, 1-24.
39. Kumar, A. V., Garg, V. K., & Buttar, H. S. (2025). Harnessing CRISPR/Cas systems for tailored therapeutic interventions in molecular medicine: Advancements in precision medicine and enhanced patient care. In *Molecular Medicine and Biomedical Research in the Era of Precision Medicine* (pp. 397-425). Academic Press.
40. Cabré-Romans, J. J., & Cuella-Martin, R. (2025). CRISPR-dependent base editing as a therapeutic strategy for rare monogenic disorders. *Frontiers in Genome Editing*, 7, 1553590.
41. Levesque, S., & Bauer, D. E. (2025). CRISPR-based therapeutic genome editing for inherited blood disorders. *Nature Reviews Drug Discovery*, 1-19.
42. Shan, Y., Zhu, Z., Liu, X., Chi, L., Zhang, J., Cheng, L., & Liu, T. (2025). In-depth analysis of cystic fibrosis cases caused by CFTR gene variation and research on the prediction and simulation of the impact on protein function. *Frontiers in Pediatrics*, 13, 1574919.
43. Lemson, A., Bosteels, C., van Ingen, J., Reijers, M., Westra, D., & Hoefsloot, W. (2025). A unique combination of heterozygous CFTR gene variants in a person with cystic fibrosis and M. abscessus infection. *Respiration*.
44. Chen, G., Wei, T., Yang, H., Li, G., & Li, H. (2022). CRISPR-

- based therapeutic gene editing for Duchenne muscular dystrophy: advances, challenges and perspectives. *Cells*, 11(19), 2964.
45. Lerma, G. T., Ryhlick, K. R., Beljan, J. C., Carraher, O. M., & Amacher, S. L. (2025). Validation of Duchenne muscular dystrophy candidate modifiers using a CRISPR-Cas9-based approach in zebrafish. *bioRxiv*, 2025-05.
46. Wu, Y. F., Chen, J. A., & Jong, Y. J. (2025). Treating neuromuscular diseases: unveiling gene therapy breakthroughs and pioneering future applications. *Journal of Biomedical Science*, 32(1), 30.
47. Guruprasad, P., Ramasubramanian, R., Nason, S., Carturan, A., Liu, S., Paruzzo, L., ... & Ruella, M. (2025). Manufacturing of CRISPR-edited primary mouse CAR T cells for cancer immunotherapy. *Nature Protocols*, 1-26.
48. Lara, P., Aguilar-González, A., Martín, F., Mesas, C., Moreno, J., & Rama, A. R. (2025). Exploring miR-21 Knock-Out Using CRISPR/Cas as a Treatment for Lung Cancer. *Genes*, 16(2), 133.
49. Liu, Z., Li, Y., Wang, S., Wang, Y., Sui, M., Liu, J., ... & Hou, P. (2025). Genome-wide CRISPR screening identifies PHF8 as an effective therapeutic target for KRAS-or BRAF-mutant colorectal cancers. *Journal of Experimental & Clinical Cancer Research*, 44(1), 70.
50. Alhabbab, R. Y. (2020). Targeting cancer stem cells by genetically engineered chimeric antigen receptor T cells. *Frontiers in Genetics*, 11, 312.
51. Malik, P. (2025). Recalibrating immune balance: CRISPR based reengineering of CTLA-4 and PD-1 pathways. *JOURNAL OF IMMUNOLOGY RESEARCH*, 2(1), 1-6.
52. Villora, S. A., Zhao, Y., Silva, P. C., Hahn, A. A., Olanin, V., Groll, D., ... & Richter, A. M. (2025). Epigenetic silencing and CRISPR-mediated reactivation of tight junction protein claudin10b (CLDN10B) in renal cancer. *Clinical Epigenetics*, 17(1), 102.
53. Al-Malki, E. S. (2025). Synthetic biology and parasite genomics: engineering parasite-resistant human microbiomes for sustainable disease prevention. *Beni-Suef University Journal of Basic and Applied Sciences*, 14(1), 16.
54. Tan, X., Li, J., Cui, B., Wu, J., Toischer, K., Hasenfuß, G., & Xu, X. (2025). CRISPR/Cas13-Based Anti-RNA Viral Approaches. *Genes*, 16(8), 875.
55. Nouri, F., Alibabaei, F., Forouzanmehr, B., Tahmasebi, H., Oksenysh, V., & Eslami, M. (2025). Progress in CRISPR Technology for Antiviral Treatments: Genome Editing as a Potential Cure for Chronic Viral Infections. *Microbiology Research*, 16(5), 104.
56. Anastassopoulou, C., Tsakri, D., Panagiotopoulos, A. P., Saldari, C., Sagona, A. P., & Tsakris, A. (2025). Armed Phages: A New Weapon in the Battle Against Antimicrobial Resistance. *Viruses*, 17(7), 911.
57. Al-Ouqaili, M. T., Ahmad, A., Jwair, N. A., & Al-Marzooq, F. (2025). Harnessing bacterial immunity: CRISPR-Cas system as a versatile tool in combating pathogens and revolutionizing medicine. *Frontiers in cellular and infection microbiology*, 15, 1588446.
58. Tee, P. Y. E., Chu, S. Y. C., Kok, C. C. Y., Foo, M., Tan, C. Z. J., Foo, J. B., ... & Hamzah, S. (2025). Applications of CRISPR in Parasitology. *Current Pharmaceutical Biotechnology*.
59. Kaya, M. (2025). The Role of CRISPR Technology in Advancing Personalized Medicine: Commercial and Ethical Considerations. *Journal of Commercial Biotechnology*, 30(2), 213-223.
60. Butt, H., Mandava, M., & Jacobsohn, D. (2025). Advances in Gene Therapy for Sickle Cell Disease: From Preclinical Innovations to Clinical Implementation and Access Challenges. *The CRISPR journal*, 8(3), 174-188.
61. Otuya, D. O., Liu, Z., Joseph, R., Hanafy, M. A., Vijaykumar, K., Stanford, D., ... & Solomon, G. M. (2025). Toward in vivo bronchoscopic functional CFTR assessment using a short circuit current measurement probe. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, 328(2), L313-L320.
62. Ali, S., Arshad, M., Summer, M., Zulfiqar, M., Noor, S., Nazakat, L., & Javed, M. A. (2025). Recent developments on checkpoint inhibitors, CAR T cells, and beyond for T cell-based immunotherapeutic strategies against cancer. *Journal of Oncology Pharmacy Practice*, 10781552251324896.
63. Tolomeo, M., Tolomeo, F., & Cascio, A. (2025). The Complex Interactions Between HIV-1 and Human Host Cell Genome: From Molecular Mechanisms to Clinical Practice. *International Journal of Molecular Sciences*, 26(7), 3184.
64. Kazakova, A. A., Leonova, E. I., Sopova, J. V., Chirinskaite, A. V., Minskaya, E. S., Kukushkin, I. S., ... & Reshetnikov, V. V. (2025). Progress in CRISPR/Cas13-Mediated Suppression of Influenza A and SARS-CoV-2 Virus Infection in in vitro and in vivo Models. *Biochemistry (Moscow)*, 90(6), 786-803.
65. Kalter, N., Fuster-García, C., Silva, A., Ronco-Díaz, V.,

- Roncelli, S., Turchiano, G., ... & Hendel, A. (2025). Off-target Effects in CRISPR-Cas Genome Editing for Human Therapeutics: Progress and Challenges. *Molecular Therapy Nucleic Acids*.
66. Zhu, M., Xu, R., Yuan, J., Wang, J., Ren, X., Cong, T., ... & Lan, X. (2025). Tracking-seq reveals the heterogeneity of off-target effects in CRISPR-Cas9-mediated genome editing. *Nature Biotechnology*, 43(5), 799-810.
67. Frias, N. A. (2025). Pioneering The Development Of CRISPR-Guided Photooxidation Of Guanine For Guanine-To-Cytosine Conversion In Cellular DNA.
68. Park, S. J., Lee, G. E., Cho, S. M., & Choi, E. H. (2025). Recent applications, future perspectives, and limitations of the CRISPR-Cas system. *Molecular Therapy Nucleic Acids*.
69. Moyo, B., Brown, L. B., Khondaker, I. I., & Bao, G. (2025). Engineering adeno-associated viral vectors for CRISPR/Cas based in vivo therapeutic genome editing. *Biomaterials*, 123314.
70. Stigzelius, V., Cavallo, A. L., Chandode, R. K., & Nitsch, R. (2025). Peeling back the layers of immunogenicity in CRISPR/Cas9-based genomic medicine. *Molecular Therapy*.
71. Fang, S., Song, X., Cui, L., Hu, L., Wang, M., Ai, L., & Wang, S. (2025). Application of the *Streptococcus pyogenes* CRISPR/Cas9 system in *Lactocaseibacillus paracasei* CGMCC4691. *Journal of Future Foods*, 5(5), 520-527.
72. Seijas, A., Cora, D., Novo, M., Al-Soufi, W., Sánchez, L., & Arana, Á. J. (2025). CRISPR/Cas9 Delivery Systems to Enhance Gene Editing Efficiency. *International Journal of Molecular Sciences*, 26(9), 4420.
73. Willy, C. J., & Faria, F. N. (2025). Techno-natalism: Geopolitical and socioeconomic implications of emerging reproductive technologies in a world of sub-replacement fertility. *Politics and the Life Sciences*, 1-20.
74. Chiong, M. A. D., & Basas, A. A. (2025). Theological and Ethical Perspectives on Gene Editing and the Sanctity of Life: Rare Genetic Diseases in the Philippines as a Model. *Journal of Religion and Health*, 1-21.
75. Voigt, J. M. (2025). From concept to cure: exploring the evolution of gene therapy and gene editing for sickle cell anemia/Author Jonas M. Voigt.
76. Wei, Y., Yue, T., Wang, Y., & Yang, Y. (2025). Fertile androgenetic mice generated by targeted epigenetic editing of imprinting control regions. *Proceedings of the National Academy of Sciences*, 122(27), e2425307122.
77. Hayenhjelm, M., & Nordlund, C. (2025). The risks and ethics of human gene editing: a philosophical guide to the arguments (p. 311). Springer Nature.
78. Gurgunidze, M., & Urtmelidze, T. INTELLECTUAL PROPERTY IN GLOBAL SECURITY: BALANCING RIGHTS AND PROTECTION STRATEGIES. In *International scientific conference-BINS 2024* (p. 147).
79. Mboowa, G., Sserwadda, I., Kanyerezi, S., Tukwasibwe, S., & Kidenya, B. (2025). The dawn of a cure for sickle cell disease through CRISPR-based treatment: A critical test of equity in public health genomics. *Annals of Human Genetics*, 89(4), 188-194.
80. Moore-Igwe, B. W., & Chukwu, P. H. (2025). CRISPR-Cas9: a step toward a cure for sickle cell disease. *Bayero Journal of Medical Laboratory Science*, 9(2), 201-210.
81. Hassan, Y. M., Mohamed, A. S., Hassan, Y. M., & El-Sayed, W. M. (2025). Recent developments and future directions in point-of-care next-generation CRISPR-based rapid diagnosis. *Clinical and experimental medicine*, 25(1), 33.
82. Bansal, P., & Kaur, N. (2025). Assessing risks associated with large-scale adoption of CRISPR gene-edited crops. *Journal of Crop Science and Biotechnology*, 28(2), 155-165.
83. Nasir, R. (2025). Developing CRISPR-Based Therapies for Genetic Diseases: Clinical Trials and Regulatory Challenges. *Indus Journal of Agriculture and Biology*, 4(01), 13-26.